Acta Crystallographica Section C

# **Crystal Structure Communications**

ISSN 0108-2701

# 5-Fluoro-1-octanoyluracil

# **Lehmler and Parkin**

## **Electronic paper**

This paper is published electronically. It meets the data-validation criteria for publication in Acta Crystallographica Section C. The submission has been checked by a Section C Co-editor though the text in the 'Comments' section is the responsibility of the authors.

 $\bigcirc$  2000 International Union of Crystallography • Printed in Great Britain – all rights reserved

# electronic papers

Acta Crystallographica Section C

## Crystal Structure Communications

ISSN 0108-2701

# 5-Fluoro-1-octanoyluracil

# Hans-Joachim Lehmler<sup>a\*</sup> and Sean Parkin<sup>b</sup>

<sup>a</sup>Graduate Center for Toxicology, University of Kentucky, Lexington, KY 40536-0305, USA, and <sup>b</sup>Department of Chemistry, University of Kentucky, Lexington, KY 40506-0055, USA
Correspondence e-mail: hjlehm1@pop.uky.edu

Received 6 September 2000 Accepted 20 September 2000

Data validation number: IUC0000266

The crystal structure of 5-fluoro-1-octanoyluracil [5-fluoro-1-octanoylpyrimidine-2,4(1H,3H)-dione,  $C_{12}H_{17}FN_2O_3$ ], a lipophilic prodrug of 5-fluorouracil, is described. The 5-fluoropyrimidine-2,4(1H,3H)-dione moiety is similar to the known structure of 1-acetyl-5-fluorouracil. The 1-octanoyl group and the 5-fluorouracil moiety are essentially coplanar, with the octanoyl carbonyl group oriented towards the the ring C-H group and away from the nearer ring carbonyl group. The torsion angle C-N-C-O (from the ring CH group to the octanoyl carbonyl group) of 9.2 (2)° is similar to the corresponding torsion angles reported for 1-acetyl-5-fluorouracil (17.3 and 1.6°) and 1,3-diacetyl-5-fluorouracil (8.8°).

### Comment

The antimetabolite 5-fluorouracil is used for the treatment of solid tumors such as gastrointestinal adenocarcinoma, breast cancer and squamous cell carcinoma of the head and neck (Iyer & Ratain, 1999). 5-Fluorouracil cannot be administered orally because of its unpredictable absorption, non-linear pharmacokinetics and a high interpatient variance. Numerous 5-fluorouracil analogues have been synthesized to improve the delivery of 5-fluorouracil (Iyer & Ratain, 1999; Lamont & Schilsky, 1999; Ozaki, 1996). After delivery to the target tissue, these analogues are subject to chemical or enzymatic hydrolysis in vivo and release 5-fluorouracil (Bundgaard et al., 1983; Møllgaard et al., 1982). There is currently growing interest in lipophilic 1- and 3-acyl derivatives for transdermal drug delivery. Despite this potential application of acyl-5-fluorouracil derivatives, only the crystal structures of two acyl derivatives, namely 1-acetyl- and 1,3-diacetyl-5-fluorouracil, have been described (Beall et al., 1993).

The structures of both 1-acetyl- and 1-octanoyl-5-fluorouracil, *i.e.* the 5-fluoropyrimidine-2,4(1H,3H)-dione system, are very similar. The 1-octanoyl group and the 5-fluorouracil moiety of the title compound, (I), are essentially coplanar, with the C7=O7 carbonyl group oriented towards the C6-H group and away from the C2=O2 group. The torsion angle C6-N1-C7-O7 is 9.2 (2)° and is similar to the torsion

angles reported for 1-acetyl-5-fluorouracil (17.3 and  $1.6^{\circ}$ ) and 1,3-diacetyl-5-fluorouracil (8.8°) (Beall *et al.*, 1993). Most likely, the slight differences are due to packing effects in the crystal. Thus, the carbonyl of the 1-acyl group can be conjugated with the pyrimidine-2,4(1H,3H)-dione ring system. As a result of the orientation of the acyl group, the partially positive carbonyl C7 atom is easily accessible to nucleophiles such as hydroxide, and the hydrolysis of 1-acyl-5-fluorouracil derivatives is fast. For example, the half-life of 1-acetyl-5-fluorouracil is about 4.8 min (Beall *et al.*, 1993).

## **Experimental**

5-Fluoro-1-octanoyluracil was synthesized by acylation of 5-fluoro-uracil with octanoyl chloride (Roberts & Sloan, 1999; Taylor & Sloan, 1998). White crystals were obtained upon crystallization from diethyl ether at 253 K (m.p. 336–338 K).  $^{1}$ H NMR (CDCl<sub>3</sub>, 400 MHz): δ 0.88 (t, –CH<sub>3</sub>, J = 6.8 Hz, 3H), 1.20–1.42 (m, 8H), 1.72 (q, –CH<sub>2</sub>CH<sub>2</sub>CON, J = 7.4 Hz, 2H), 3.12 (t, –CH<sub>2</sub>CON, J = 7.4 Hz, 2H), (d, –CH=CF-, J = 6.8 Hz, 1H);  $^{13}$ C NMR (CDCl<sub>3</sub>, 75 MHz): δ 14.04 (–CH<sub>3</sub>), 22.56, 24.37, 28.84, 28.94, 31.59, 39.04, 121.75 (—CH-, J<sub>CF</sub> = 27 Hz), 141.26 (—CF-, J<sub>CF</sub> = 182 Hz), 147.75 (N – CO – NH), 156.62 (—CF – CO – NH, J<sub>CF</sub> = 21 Hz), 171.98 (acyl CO);  $^{19}$ F NMR (CDCl<sub>3</sub>): δ –161.10 (d, 6.0 Hz); IR (cm<sup>-1</sup>): 1738 and 1704 [ $\nu$ (C=O)]; MS m/z (relative intensity, %): 256 (1, M<sup>+</sup>), 127 (100, C<sub>8</sub>H<sub>15</sub>O<sup>+</sup>), 57 (84), 43 (31).

### Crystal data

- 2	
$C_{12}H_{17}FN_2O_3$	$D_x = 1.347 \text{ Mg m}^{-3}$
$M_r = 256.28$	Mo $K\alpha$ radiation
Triclinic, $P\overline{1}$	Cell parameters from 4236
a = 5.4500 (11) Å	reflections
b = 9.7410 (19) Å	$\theta = 1.00 - 25.35^{\circ}$
c = 12.307 (3)  Å	$\mu = 0.107 \text{ mm}^{-1}$
$\alpha = 80.27 (3)^{\circ}$	T = 173 (1)  K
$\beta = 85.97 (3)^{\circ}$	Irregular plate-like
$\gamma = 79.13 (3)^{\circ}$	fragment, colourless
$V = 631.9 (2) \text{ Å}^3$	$0.32 \times 0.20 \times 0.04 \text{ mm}$
Z = 2	

#### Data collection

Nonius KappaCCD diffractometer	$\theta_{\rm max} = 25.23^{\circ}$
$\omega$ scans at fixed $\chi = 55^{\circ}$	$h = -6 \rightarrow 6$
4358 measured reflections	$k = -11 \rightarrow 11$
2277 independent reflections	$l = -14 \rightarrow 14$
1562 reflections with $I > 2\sigma(I)$	Intensity decay: <1%
$R_{\rm int} = 0.034$	

#### Refinement

Refinement on $F^2$	$w = 1/[\sigma^2(F_o^2) + (0.0424P)^2]$
$R[F^2 > 2\sigma(F^2)] = 0.041$	where $P = (F_o^2 + 2F_c^2)/3$
$wR(F^2) = 0.100$	$(\Delta/\sigma)_{\rm max} < 0.001$
S = 1.046	$\Delta \rho_{\text{max}} = 0.21 \text{ e Å}^{-3}$
2277 reflections	$\Delta \rho_{\min} = -0.20 \text{ e Å}^{-3}$
165 parameters	Extinction correction: SHELXL97
H-atom parameters constrained	Extinction coefficient: 0.048 (5)

Table 1 Selected geometric parameters ( $\mathring{A}$ ,  $^{\circ}$ ).

N1-C6	1.3950 (19)	C5-F5	1.3490 (18)
N1-C2	1.410(2)	O7-C7	1.2084 (18)
N1-C7	1.449 (2)	C7-C8	1.500(2)
C2-O2	1.2074 (18)	C8-C9	1.527 (2)
C2-N3	1.3766 (19)	C9-C10	1.517 (2)
N3-C4	1.3714 (19)	C10-C11	1.523 (2)
C4-O4	1.2282 (18)	C11-C12	1.520(2)
C4-C5	1.440(2)	C12-C13	1.514(2)
C5-C6	1.316(2)	C13-C14	1.517(2)
C6-N1-C2	120.36 (14)	F5-C5-C4	116.15 (14)
C6-N1-C7	115.90 (13)	C5-C6-N1	121.15 (15)
C2-N1-C7	123.71 (13)	O7 - C7 - N1	116.73 (14)
O2-C2-N3	121.57 (14)	O7 - C7 - C8	123.48 (15)
O2-C2-N1	124.02 (15)	N1-C7-C8	119.78 (14)
N3-C2-N1	114.41 (14)	C7-C8-C9	111.77 (14)
C4-N3-C2	128.42 (14)	C10-C9-C8	112.19 (13)
O4-C4-N3	122.42 (15)	C9-C10-C11	114.19 (14)
O4-C4-C5	125.06 (16)	C12-C11-C10	113.52 (14)
N3-C4-C5	112.53 (15)	C13-C12-C11	114.28 (14)
C6-C5-F5	120.89 (14)	C12-C13-C14	113.15 (14)
C6-C5-C4	122.95 (16)		

Data collection: *COLLECT* (Nonius, 1998); cell refinement: *SCALEPACK* (Otwinowski & Minor, 1997); data reduction: *DENZO–SMN* (Otwinowski & Minor, 1997); program(s) used to solve structure: *SHELXS*97 (Sheldrick, 1997); program(s) used to refine structure: *SHELXL*97 (Sheldrick, 1997); software used to

prepare material for publication: SHELXL97 (Sheldrick, 1997) and local programs.

HJL would like to thank Dr L. W. Robertson for providing support and laboratory facilities and Dr J. Goodman from the University of Kentucky Life Sciences Mass Spectrometry Facility for performing the mass spectral analysis. This work was supported in part by the University of Kentucky Medical Center Research Fund.

#### References

Beall, H. D., Prankerd, R. J., Todaro, L. J. & Sloan, K. B. (1993). Pharm. Res. 10, 905–912.

Bundgaard, H., Hoelgaard, A. & Møllgaard, B. (1983). *Int. J. Pharm.* **15**, 285–292.

Iyer, L. & Ratain, M. J. (1999). Cancer Invest. 17, 494-506.

Lamont, E. B. & Schilsky, R. L. (1999). Clin. Cancer Res. 5, 2289–2296.
 Møllgaard, B., Hoelgaard, A. & Bundgaard, H. (1982). Int. J. Pharm. 12, 153–162

Nonius (1998). *COLLECT*. Nonius BV, Delft, The Netherlands. Otwinowski, Z. & Minor, W. (1997). *Methods Enzymol.* **276**, 307–326. Ozaki, S. (1996). *Med. Res. Rev.* **16**, 51–86.

Roberts, W. J. & Sloan, K. B. (1999). J. Pharm. Sci. 88, 515-522.

Sheldrick, G. M. (1997). SHELXL97 and SHELXS97. University of Göttingen, Germany.

Taylor, H. E. & Sloan, K. B. (1998). J. Pharm. Sci. 87, 15-20.